

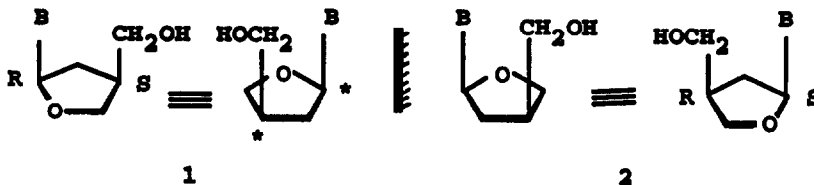
NOVEL ISOMERIC DIDEOXYNUCLEOSIDES OF THE D- AND L-APIOSE FAMILY

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Summary: Short synthetic approaches to optically active, *cis* and *trans* dideoxynucleoside analogs of the D- and L-ribose family have been developed. The chiral precursor for the syntheses was the enzymatically prepared compound, S(-)-2-(2-propenyl)-1,3-propanediol monoacetate (5).

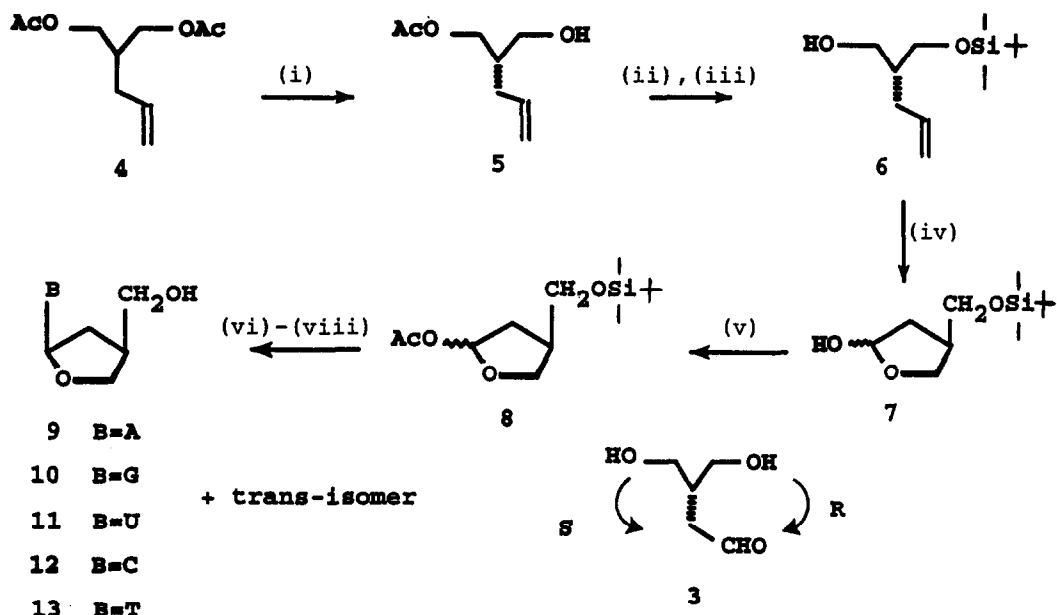
A few dideoxynucleosides, through inhibition of HIV-encoded reverse transcriptase (RT), have proven to be effective pro-drugs for clinical use in the treatment of AIDS.¹ The design and evaluation of additional novel nucleoside-based RT inhibitors are needed in order to develop analogs that exhibit a more favorable toxicity profile and are less susceptible to the development of resistant strains of HIV.² One essential feature in the design of these inhibitors is retention of the 2',3'-dideoxylation which is necessary for termination of the viral DNA chain elongation. The most common modification of dideoxynucleosides has been strategic substitution on the carbohydrate moiety (i.e. azido and fluoro).^{3,4} A more recent trend in design has been antiviral dideoxynucleosides with no heteroatom or an additional heteroatom within the carbohydrate moiety.⁵⁻¹⁰

An alternative approach involves dideoxygenated nucleosides that contain transposed heteroatoms¹¹⁻¹³ and are regioisomeric with respect to dideoxy analogs of the natural nucleosides. 9-(β-D-Apio-D-furanosyl)adenine, a biologically-active, relatively non-cytotoxic nucleoside¹⁴⁻¹⁷ related to natural D-ribose,¹⁸ is a regioisomer of adenosine through transposition of the C-4' hydroxymethyl to C-3'. We wish to report on the stereoselective synthesis of the complete family of 2',3'-dideoxygenated nucleosides (1 and 2) related to apio nucleosides as potential inhibitors of HIV replication. This study is supported by the observation¹³ that one member of Class 2 has been reported to have anti-HIV activity in MT-4 cells with no apparent toxicity.



The key precursor for the construction of the dideoxyapiose ring was a derivative of the optically pure aldodiol system 3, the cyclization of which in one direction creates the carbon bearing the CH₂OH of R-stereochemistry and in the other direction of S-stereochemistry. This approach allows a shorter synthetic route

than one involving D- or L-apiose and would avoid potential problems such as racemization associated with the deoxygenation of the C-3 tertiary hydroxyl group of apiose. The starting compound for the enantioselective step to the chiral precursor **5**^{19,20} was 2-(2-propenyl)-1,3-propanediol diacetate (**4**), prepared in two steps (reduction followed by acetylation) from diethyl allylmalonate in 80% overall yield (Scheme 1). Stereoselective deacetylation with the lipase from *Candida cylindracea* (Sigma, Type VII) afforded the S-(-)-monoacetate of 2-(2-propenyl)-1,3-propanediol (**5**) ($[\alpha]_D = -8.0^\circ$, CHCl₃) in a 50% yield (99% ee). Treatment of **5** with *t*-butyldimethylsilyl chloride followed by deesterification gave the R-(+)-**6** ($[\alpha]_D = +3.7^\circ$, CHCl₃) in 96% overall yield for the two steps. For the key transformation, the formation of the 2,3-dideoxy-D-ribofuranosyl system from **6**, a variety of conditions were examined. The most successful method was the oxidative cleavage of the olefin employing OsO₄/NaIO₄. Thus, treatment of **6** with OsO₄ and NaIO₄ provided **7** almost quantitatively as an anomeric mixture ($[\alpha]_D = +24^\circ$, CHCl₃) which, upon acetylation, gave the corresponding 1-O-acetyl-3'-O-(*t*-butyldimethylsilyl)-2,3-dideoxy-D-ribose (**8**) in a 78% yield. Trimethylsilyl triflate promoted condensation²¹



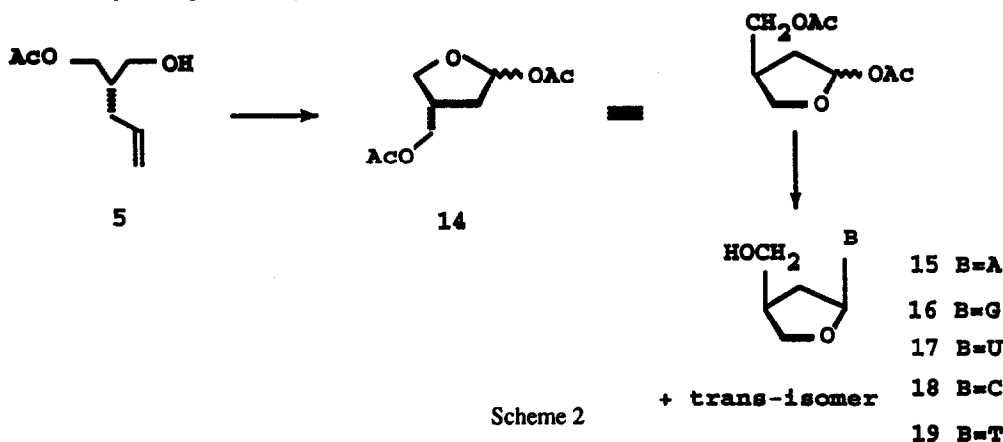
(i) Lipase (0.4 g per mmol **4**), 30% aq. acetone, NaOH (1M, pH 7), 8 h; (ii) TBDMSiCl (1.2 eq), imidazole (1.2 eq), CH₂Cl₂, 24 h; (iii) NaOMe (1.2 eq), MeOH, 45 min; (iv) OsO₄ (0.05 eq), NaIO₄ (3 eq), H₂O/Et₂O (50% v/v), 12 h; (v) Ac₂O (1.2 eq), Et₃N (1.4 eq), DMAP (0.1 eq), CH₂CN, 1 h; (vi) purine or pyrimidine base (1.2 eq), bis(trimethylsilyl)acetamide (1.3-2.6 eq), CH₂CN, 82 °C; (vii) TMSOTf (1.1 eq), 0 °C - R.T., 2-5 h; (viii) NH₃/MeOH and/or Et₃NF (2 eq).

Scheme 1

of **8** with silylated N⁶-benzoyladenine, generated *in situ*, gave a 3:2 (α : β) diastereomeric mixture of anomeric adenine isodideoxynucleosides in 43% yield. Separation by preparative layer chromatography and quantitative deprotection of the individual anomers provided 9-(2,3-dideoxy- β -D-apiofuranosyl)adenine (**9**) ($[\alpha]_D = -22.6^\circ$, MeOH) and the corresponding 9-(2,3-dideoxy- α -D-apiofuranosyl)adenine anomer ($[\alpha]_D = +39.8^\circ$, MeOH). Assignments of the anomeric configurations were readily determined through ¹H NMR NOE difference spectroscopy.

Under similar conditions, the persilylated bases of N²-acetyl-O⁶-diphenylcarbamoylguanine, uracil, cytosine, and thymine were coupled with the acetylated dideoxyapiose **8**. Separation of the resulting α and β anomers and deprotection gave the 2',3'-dideoxy- β -D-apiofuranosyl nucleosides **10-13**. In the case of the cytosine and thymine apiosyl nucleosides, the separation is more laborious due to a small difference only in R_f values between the anomers.

The nucleosides of the 2,3-dideoxy-L-apiofuranosyl series (Scheme 2) were similarly obtained from the chiral precursor **5**. When **5** was treated with OsO₄/NaIO₄, followed by acetylation, **14** was formed in 63% yield. Glycosylation with the appropriate silylated aglycon, diastereoisomer separation, and deprotection provided the 2',3'-dideoxy- α -L-apiofuranosyl nucleosides **15-19**.



The compounds of these apio dideoxynucleoside families are resistant to enzymatic deamination (e.g. the substrate activity of **9** towards mammalian adenosine deaminase is 0.12% compared to adenosine. Studies of the relative rates of glycosidic bond hydrolysis²² show that these compounds are slightly more stable than 2',3'-dideoxynucleosides (e.g. compound **9** is hydrolyzed at 84% of the rate of 2',3'-dideoxyadenosine at pH 3). Comprehensive antiviral studies are currently in progress and those results will be reported elsewhere.

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